Development of Protein Polycrystallography

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Introduction – powders?
Real time & radiation damage studies

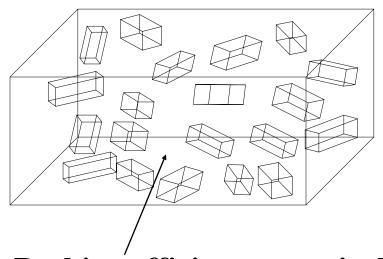
Ab initio??

References:

Modern Powder Diffraction, Eds. J. Post & D. Bish The Rietveld Method, Ed. R.A. Young Von Dreele, J. Appl. Cryst. 32, 1084 (1999), Von Dreele, et al., Acta Cryst. D56, 1549-1553 (2000), Von Dreele, Acta Cryst. D57, 1836-1842 (2001)



What is a powder? - polycrystalline mass



All orientations of crystallites possible

Sample: $1\mu l$ powder of $1\mu m$ crystallites - $\sim 10^9$ particles

Packing efficiency – typically 50% Spaces – air, solvent, etc.

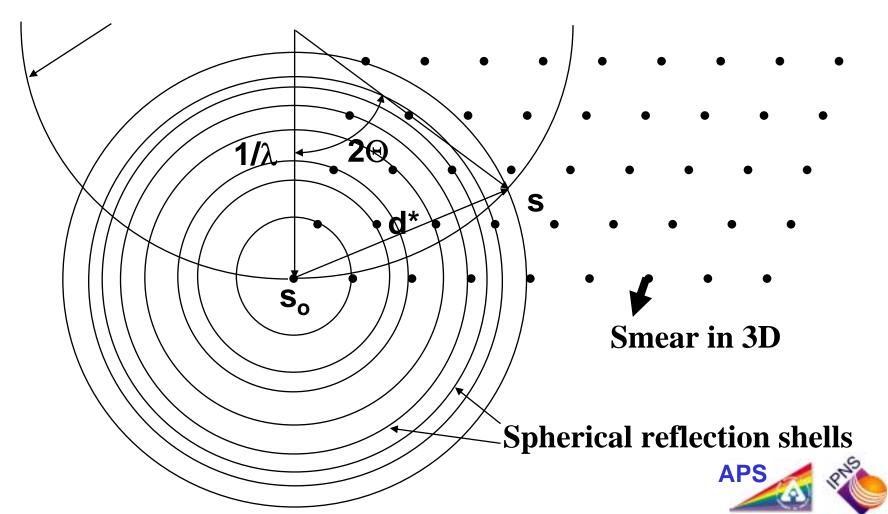
Single crystal reciprocal lattice
- smeared into spherical shells



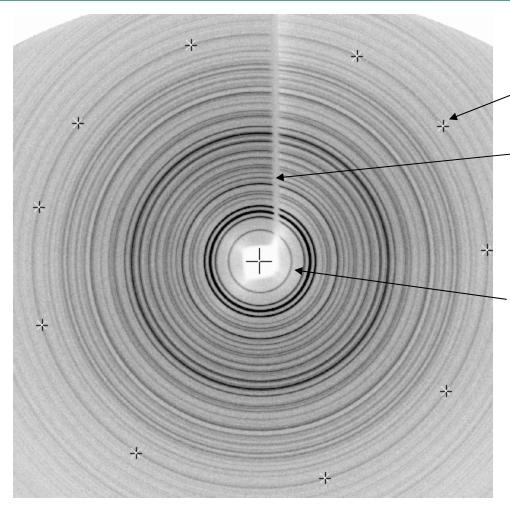
Powder diffraction - reciprocal space

Bragg's Law $d*/2 = \sin \Theta/\lambda$

Ewald sphere



Rings – protein pattern (HEWL)



Centering ring @ 4.6deg 2Θ

Beam stop holder

Inner most ring – d~55Å (110) Reflection, lowest order for tetragonal lysozyme 2Θ ~ 0.67deg

Texture free sample & no graininess



Rietveld refinement – extract structure factors

Rietveld Minimize

$$M_R = \sum w(I_o - I_c)^2$$

Apportion I_o by ratio of I_c to ΣI_c & apply corrections

$$|F_o|^2 = \frac{1}{Lp} \sum I_o \left(\frac{I_c}{\sum I_c} \right)$$

Exact overlaps

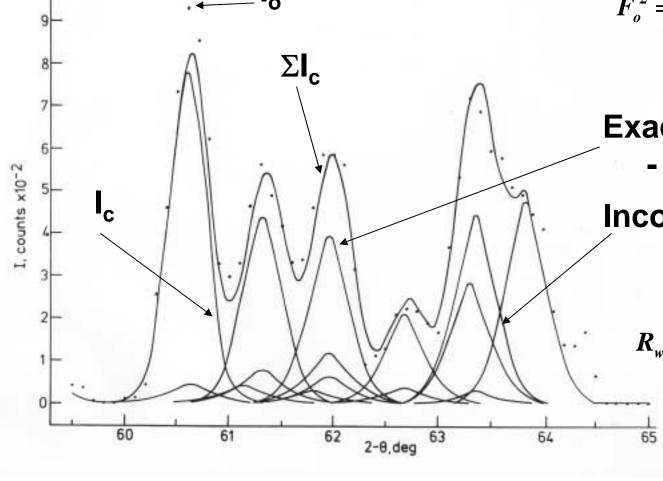
- symmetry

Incomplete overlaps

Residuals:

$$R_{wp} = \sqrt{\frac{\sum w(I_o - I_c)^2}{\sum wI_o^2}}$$





Stereochemically restrained Rietveld refinement - minimization function (typical protein technique)

$$M = f_y \sum w_i (Y_{oi} - Y_{ci})^2$$
 Powder profile (Rietveld)
 $+ f_a \sum w_i (a_{oi} - a_{ci})^2$ Bond angles
 $+ f_d \sum w_i (d_{oi} - d_{ci})^2$ Bond distances
 $+ f_t \sum w_i (-T_{ci})^2$ Torsion angle pseudopotential
 $+ f_p \sum w_i (-p_{ci})^2$ Plane RMS displacements
 $+ f_v \sum w_i (v_{oi} - v_{ci})^4$ van der Waals distances
 $+ f_h \sum w_i (h_{oi} - h_{ci})^2$ Hydrogen bonds
 $+ f_x \sum w_i (x_{oi} - x_{ci})^2$ Chiral volumes
 $+ f_R \sum w_i (-R_{ci})^4$ " ϕ/ψ or χ_1/χ_2 " pseudopotential

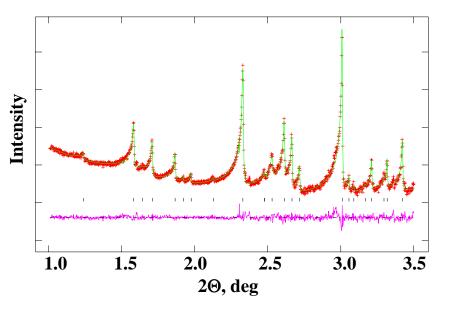
 $w_i = 1/\sigma^2$ weighting factor

 f_x - weight multipliers (typically 0.1-10)

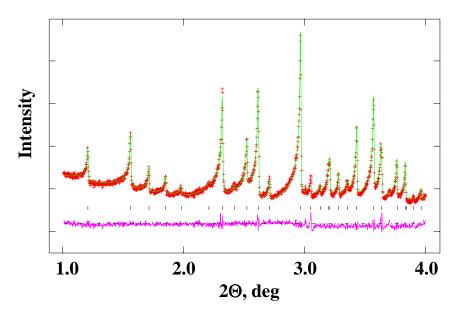


T₃R₃ Zn insulin – phase change by grinding

Grind T_3R_3 complex in agate mortar with mother liquor High resolution synchrotron x-ray powder patterns (X3b1/NSLS)



Immediately after grinding Indexed – R3 $a=81.275\text{\AA}$, c=73.024Å New phase – T_3R_3DC Solve by molec. repl.



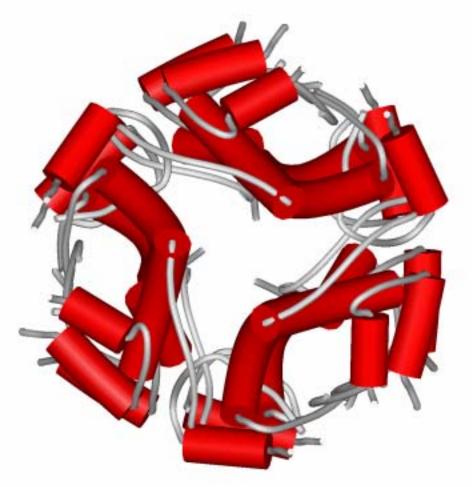
After 2 days rest Indexed – R3 a=81.084Å, c=37.537Å same as single xtal

APS

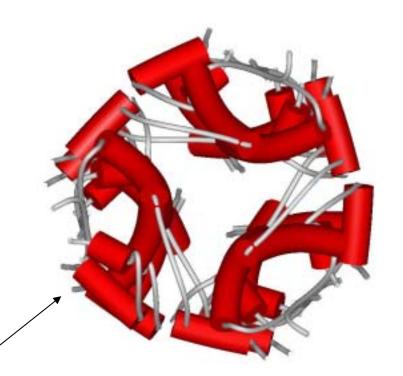
Doubled cell

Schematic of T₃R₃DC Zn-insulin complex.

Powder RT structure PDB=1FUB



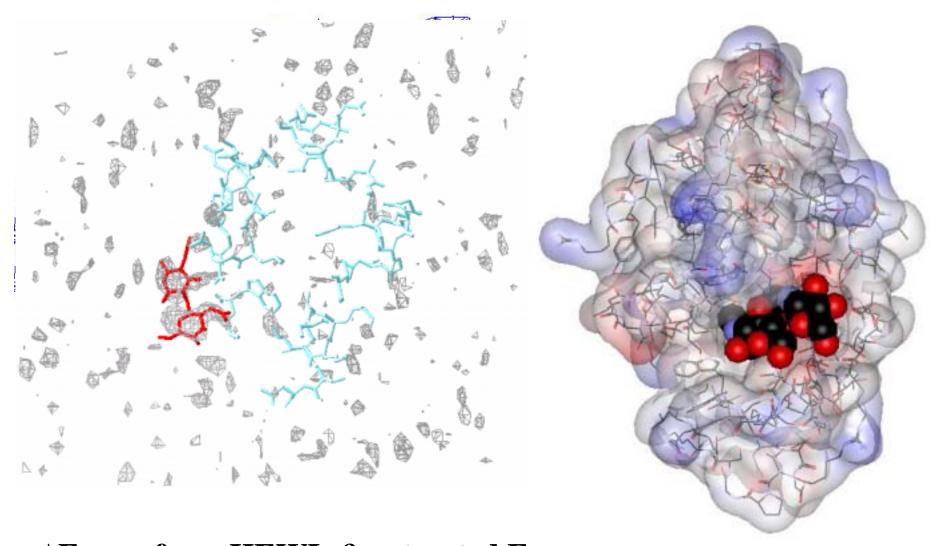
View down 3-fold axis Front T_3R_3 turned 9° wrt back T_3R_3



Same structure as -Single crystal – Lo T phase PDB=1G7A



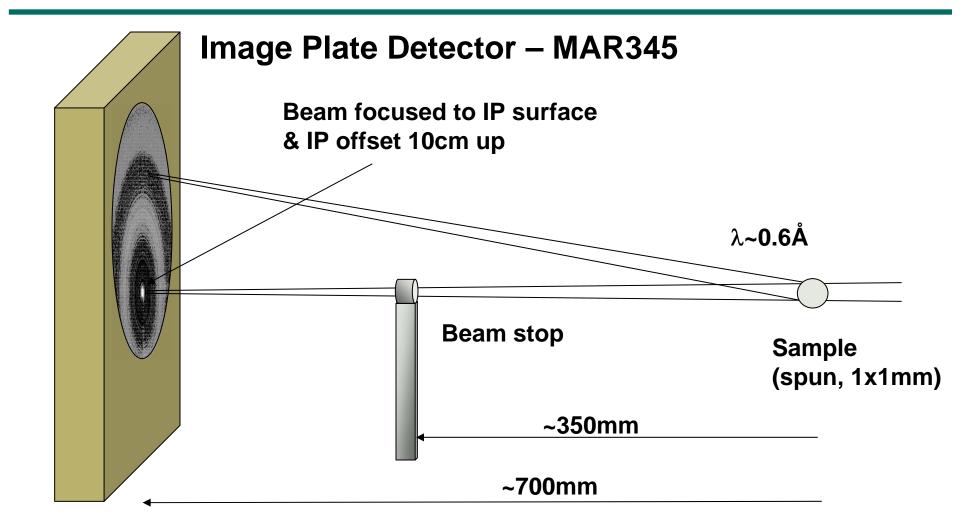
$LYSO/NAG_2$ – powder data from complex



 ΔF map from HEWL & extracted F_o



Better data collection

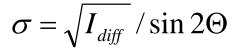




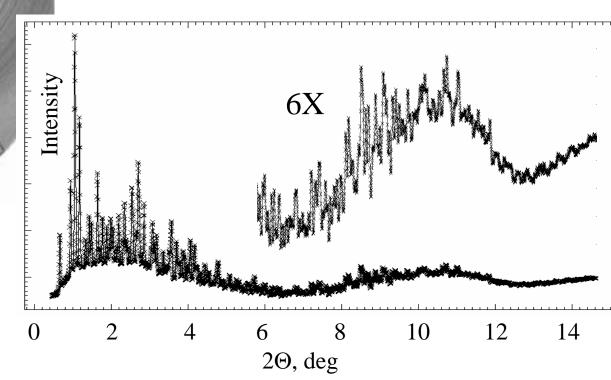
Integration – fit2d "cake" option

~60°

 d_{min} <2.5Å



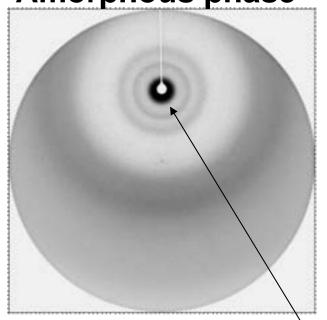
Includes effect of increase in pixels with angle



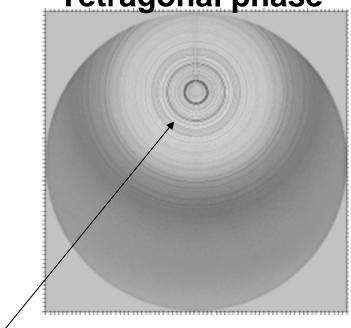


Lysozyme: 1BM; L=724mm; λ =0.6194Å; 30sec images

1.5M NaCl/ph5 buffer Amorphous phase



4.5hrs later (new sample)
Tetragonal phase

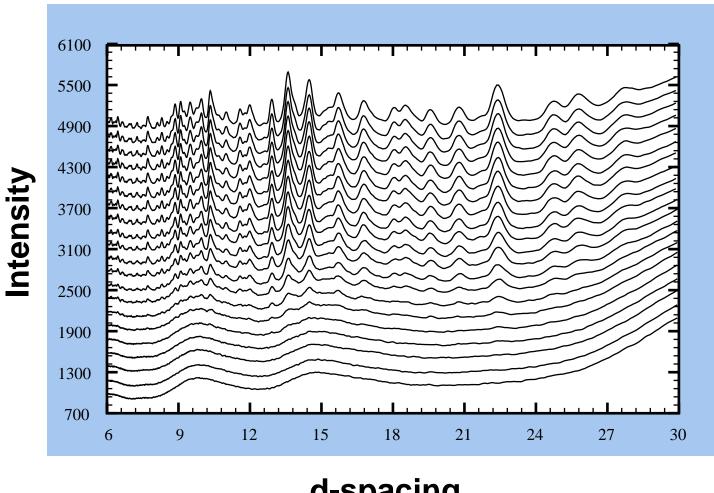


Small angle scattering & diffuse scattering

Do time series experiment & watch crystals form



Protein crystallogenisis from amorphous phase



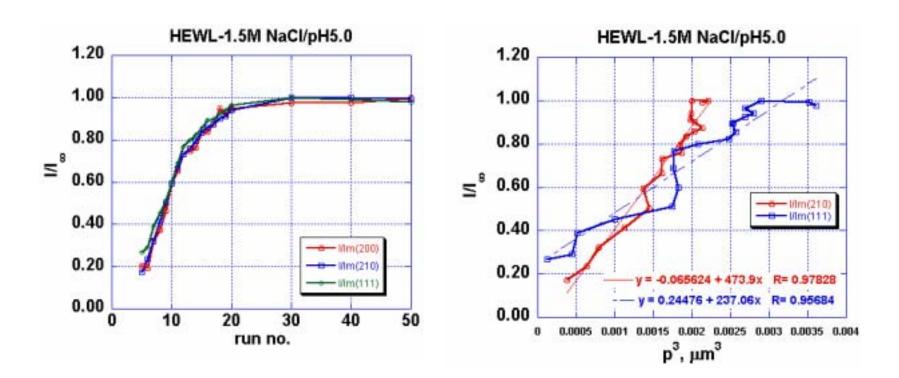
Time series:
5min steps
15min from
mixing
Results:
All growth
Vary lattice
500Å-1µm
xtals
Rad. inhibit
nucleation

d-spacing





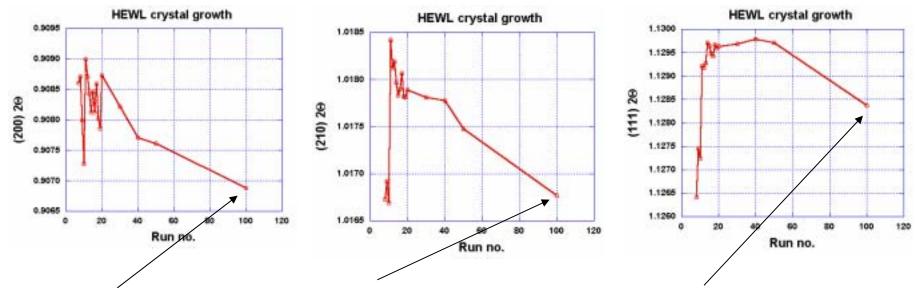
HEWL crystal growth – intensity vs particle size



All growth – no nucleation – radiation inhibited Tricky expts.



HEWL crystal growth – surface tension effects?

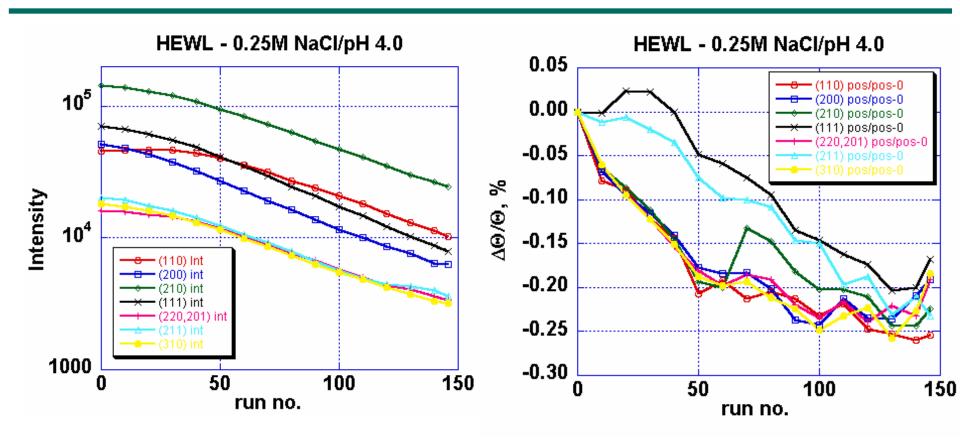


Last point (Run 100) – fresh sample complete crystallization Grain size $500\text{\AA} \rightarrow 0.8 \mu\text{m}$ Lattice decrease first & then slow increase?

Surface tension effect? ∆a~0.2Å



Radiation damage – reflection intensities & positions



APS 1BM - 30s exposures + 150s delay, room temp.
Full sequence wrt NaCl & pH – effects??
Immediate changes seen
2 stages? - <20min & >20min exposure



Protein polycrystallography – status?

Fast data collection – image plates; 30s on 1BM;

0.035° 2⊕ peaks; ~2Å

Track crystallization; radiation damage, etc.

Wide range of conditions (solvent, temp., etc.) accessible

Data analysis – restrained Rietveld refinement

Refine from starting models

Molecular replacement structure solution

"Heavy molecule" solution – complexes

1 trial of Xe adsorption – success??



Protein Polycrystallography – future?

Data collection – higher resolution IP (<100μm vs 300μm); peaks at "best" possible width (~0.01° 2Θ) to <2Å; lower background; <10s exposure with fast readout. Faster *in situ* studies (<10s compared to 150s each step) Refinement techniques – use more SC ideas *Ab initio* structure solution – multiple data extraction; heavy atom methods, etc.

